

Effective potential studies for some new hybrid molecules for their activity against prostate cancer

Estudios efectivos de potencial para algunas nuevas moléculas híbridas para su actividad contra el cáncer de próstata

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ABSTRACT

Objective: The present work aimed at developing novel hybrid molecules for targeting the prostate cancer. It is observed that two human shock proteins Hsp70 and Hsp90 are over-expressed in prostate cancer making them one of the important drug targets. We have designed and developed twelve new hybrid molecules 6a-j for targeting these proteins.

Methods: The designed molecules were prepared following a four step reaction protocol and characterized on the basis of proton NMR and Mass spectrometry. These were subjected to in vitro studies by means of Oncotest and CCK-8 assays with two cell lines DU145 and 22Rv1. The selected molecules 6b and 6i were subjected to molecular docking and then for SPR based affinity assay.

Results: Compounds 6b and 6i were found to be highly active anticancer compounds comparable to standard drug enzalutamide. They have significant IC_{50} and high dock score for the Hsp70 and Hsp90. These compounds are selective and have good binding affinity for the Hsp70 due to high Kd.

Conclusion: Compound 6b and 6i can serve as lead molecules for the development of antiprostata cancer drugs with Hsp70 as target.

Keywords: Hybrid; DU145; 22Rv1; SPR; Molecular docking.

RESUMEN

Objetivo: El presente trabajo tuvo como objetivo desarrollar nuevas moléculas híbridas para atacar el cáncer de próstata. Se observa que dos proteínas de choque humano, Hsp70 y Hsp90, se sobreexpresan en el cáncer de próstata, lo que las convierte en uno de los objetivos farmacológicos importantes. Hemos diseñado y desarrollado doce nuevas moléculas híbridas 6a-j para dirigir estas proteínas.

Métodos: Las moléculas diseñadas se prepararon siguiendo un protocolo de reacción de cuatro etapas y se caracterizaron sobre la base de RMN de protón y espectrometría de masas. Estos se sometieron a estudios in vitro por medio de ensayos Oncotest y CCK-8 con dos líneas celulares DU145 y 22Rv1. Las moléculas seleccionadas 6b y 6i se sometieron a acoplamiento molecular y luego a ensayo de afinidad basado en SPR.

Resultados: Se descubrió que los Compuestos 6b y 6i son compuestos anticancerígenos muy activos comparables al fármaco estándar enzalutamida. Tienen un IC_{50} significativo y una puntuación alta para el muelle de Hsp70 y Hsp90. Estos compuestos son selectivos y tienen una buena afinidad de unión por la Hsp70 debido a la alta Kd.

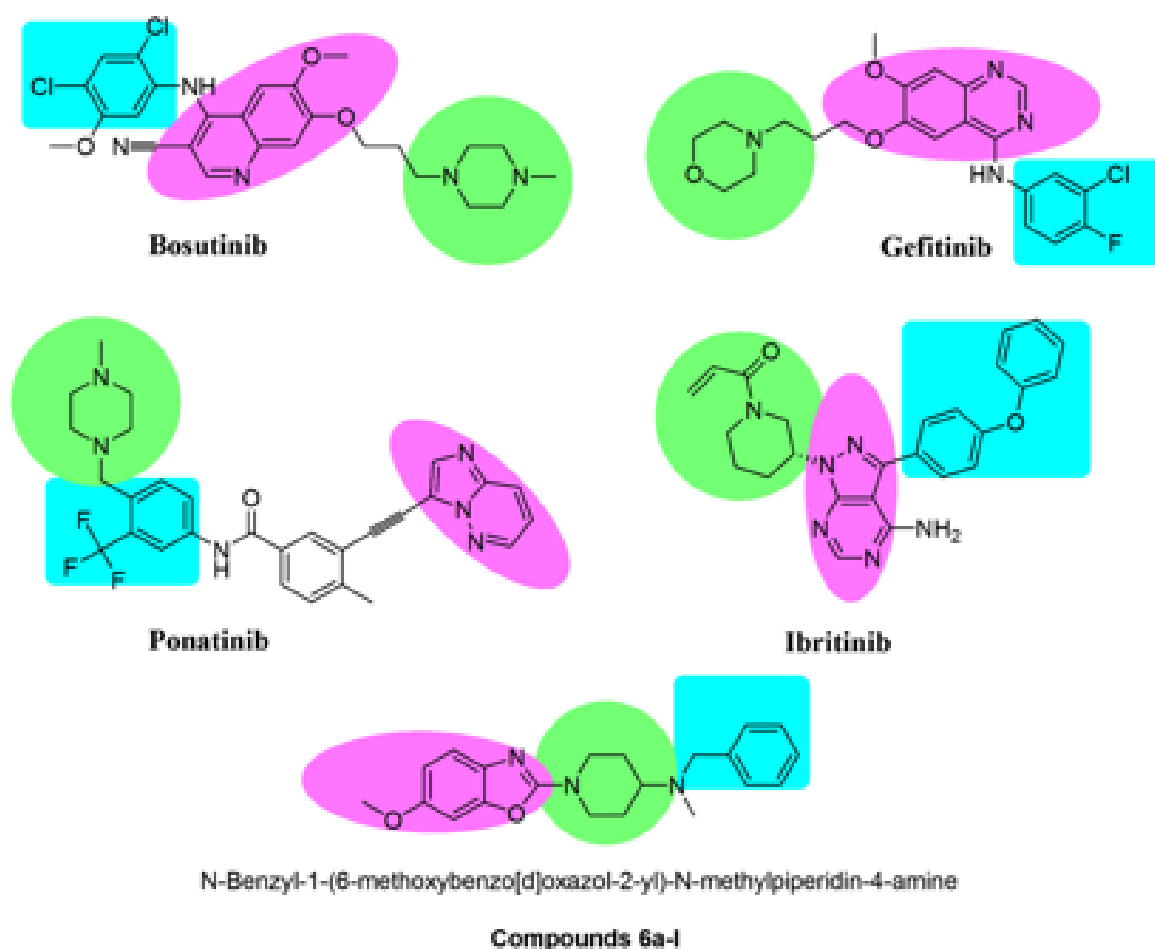
Conclusión: Los compuestos 6b y 6i pueden servir como moléculas principales para el desarrollo de fármacos antiprostáticos contra el cáncer con Hsp70 como objetivo.

Palabras clave: Híbrido; DU145; 22Rv1; SPR; acoplamiento molecular.

INTRODUCTION

The heat shock proteins (Hsp) are induced in response to heat shock and other conditions such as chemical or physical stress conditions; these are encoded by the Hsp 27, 28, 40, 70, 90 and 110 genes. These proteins possess the property of modifying other proteins, their functions and these interact to alter their normal functioning. They are highly efficient in synthesis of proteins in response to stress and have highly efficient stabilisation effect thus maintaining the normal homeostasis by transcriptional activation, mRNA stabilisation and translation.¹ Several studies have proved the role of Hsp70 and 90 in various biochemical and physiological process such as regulation of apoptosis via the apoptotic pathway, regulation for release and transport of mitochondrial cytochrome c, regulating the tumour necrosis factor (TNF) and its related apoptotic moieties to respective receptors.² Hsp 70 and 90 are highly expressed in cancer that are

drug resistant and are often observed in patients suffering from drug resistance to chemotherapeutic agents.³ Thus, these proteins have become logical and popular targets for development of anticancer agents. On this front very few drugs are available with Geldanamycin, Macbecin and Radicolas only compounds available for inhibiting the Hsp90. Whereas, Apoptozole, dibenzyladenosine analogues, Pifithrin, several pyrimidine analogues and flavonoids are found to inhibit Hsp70, but all these compounds are under investigation or in clinical phase.⁴ It has been well observed that overexpression of these heat shock proteins is one of the factors in progression of prostate cancer. Thus development of compounds that can inhibit these proteins could possibly lead to control over prostatic adenocarcinoma.^{5,6} This situation inspired us for development of some newer derivatives as inhibitors of Hsp70 and Hsp90 for treatment of prostate cancer.

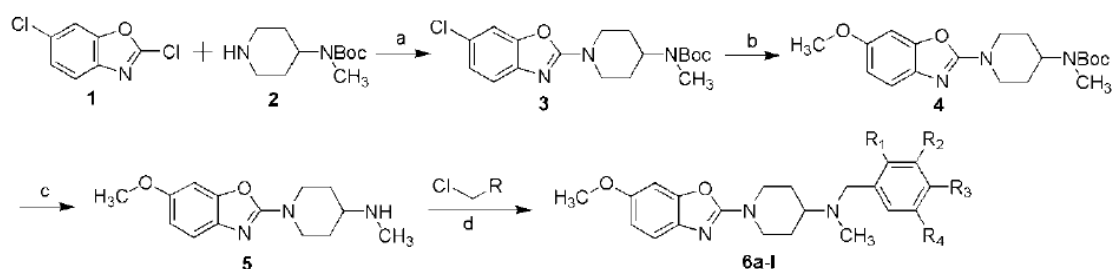


* Green Circle: Cyclic aliphatic group
 Pink Circle: Fused aromatic nucleus,
 Aquamarine colour: Aromatic substitution with halogens

Figure 1: Anticancer drugs with hybrid nature and possess common three pharmacophore features as cyclic aliphatic group with substitution, fused aromatic nucleus and the aryl substitutions generally acting as linker.

Several anticancer compounds are reported to have inhibitory activity towards the Hsp90 and Hsp70. On analysing these compounds it was found that these molecules have several common structural features and belongs to the 'nib' class of drugs. Compounds such as Bosutinib, Gefitinib, Ponatinib, Imatinib, Ceitinib, Afatinib and Ibrutinib have inhibitory action on the Hsp90 and 70.⁷⁻¹⁰ It was observed that these compounds have several common structural features such as fused ring structures or have an aliphatic cyclic moiety with heteroatom and substituted aryl ring (fig.1). Along with these structural features it was observed

that these compounds are hybrid in nature, formed from two or more biologically active chemical moieties. Benzoxazole are reported to possess antitumor activity and can be synthesised by various routes, similarly piperidine is also reported to have anticancer activity.¹¹⁻¹³ Based on these observations we designed novel hybrid molecules of benzoxazole and piperidine with structural features similar to the cancer therapeutic agents. Benzoxazole was linked with substituted piperidine to yield twelve derivatives with different substitutions on the aryl ring (Scheme 1).



Compd no.	R ₁	R ₂	R ₃	R ₄
6a	H	H	H	H
6b	H	H	Cl	H
6c	H	H	Br	H
6d	H	H	F	H
6e	H	Cl	Cl	H
6f	H	H	CF ₃	H
6g	H	H	CH ₃	H
6h	H	H	CH ₃ (CH ₂) ₃	H
6i	H	H	NO ₂	H
6j	CH ₃	H	H	CH ₃
6k	OCH ₃	H	H	OCH ₃
6l	H	OCH ₃	OCH ₃	OCH ₃

Scheme 1. Reagents and conditions for the synthesis of **6a-l**: a) Dimethyl formamide (DMF), Potassium carbonate (K₂CO₃), room temperature, 6 h; b) Sodium methoxide (CH₃ONa), Methanol, reflux, 12 h; c) Potassium Iodide (KI), Tetrahydrofuran (THF), Trifluoroacetic acid, DCM, room temperature, 7 h; d) K₂CO₃, DMF, 4 h.

MATERIAL AND METHODS

Chemicals and solvents for this work were obtained from Alfa Aesar, UK and Sigma Aldrich, USA unless otherwise mentioned. Thiele's tube melting-point apparatus was used to determine the melting points (mp) for all the compounds within open capillaries and was recorded without corrections. Infrared (IR) spectra were developed and created on JASCO FT/IR-4000 spectrophotometer (Jasco, USA) with KBr pellets used for compounds. Proton NMR (^1H) spectra were recorded on a Bruker Advance-II 400 Spectrometer on 400 MHz using tetramethylsilane (TMS) as internal standard. Chemical shift values for the synthetic derivatives were recorded as δ (parts per million), coupling constant value J was measured in hertz (Hz). The peaks are presented as s = singlet, d = doublet, t = triplet, dd = double doublet, m = multiplet. In similar way ^{13}C NMR were recorded. Thin Layer Chromatographic (TLC) method was used to determine the purity of compounds and progress of reaction using Merck, silica gel, HF254–361, type 60, 0.25 mm. Electron Spray Ionisation Mass Spectrometry (ESI-MS) were recorded on Waters HPLC system with Q-Time of Flight LC-MS spectrometry (Waters-Micromass LC-MS, US).

Chemistry

The title compounds N-substituted benzyl-1-(6-methoxybenzo[d]oxazol-2-yl)-N-methylpiperidin-4-amine **6a-l** were obtained starting from benzoxazole in four step synthetic procedure, these procedures are discussed below as follows;

Synthesis of tert-butyl 1-(6-chlorobenzo[d]oxazol-2-yl)piperidin-4-yl(methyl)carbamate 3

A mixture of 2,6-dichlorobenzo[d]oxazole **1** (1.0 mmol) and tert-butyl methyl(piperidin-4-yl)carbamate **2** (1.0 mmol) were stirred in DMF (5 mL) in presence of potassium carbonate for about 6 hours at room temperature, completion of reaction was monitored on TLC. The resulting compound tert-butyl 1-(6-chlorobenzo[d]oxazol-2-yl)piperidin-4-yl(methyl) carbamate **3** was isolated as off white ppt in 77% yield. Mp: 230-232 °C; ^1H -NMR (CDCl_3): δ 1.49 (s, 9H, Boc-CH₃), 1.60 (m, 2H, Piperidine-H), 1.89 (m, 2H, Piperidine-H), 2.3 (m, 2H, Piperidine-H), 2.5 (m, 2H, Piperidine-H), 2.9 (s, 3H, N-CH₃), 3.72 (m, 2H, methylene-H), 7.20 (m, 1H, Ar-H), 7.85 (m, 1H, Ar-H); Anal. Calcd for C₁₈H₂₄ClN₃O₃; C, 59.09; H, 6.61; Cl, 9.69; N, 11.49; O, 13.12; MS (ESI) m/z: 365.1506.

Synthesis of tert-butyl 1-(6-methoxybenzo[d]oxazol-2-yl)piperidin-4-yl(methyl)carbamate 4

Compound **3** (10 mmol) was taken in methanol (100 mL) and allowed to solubilise, to this was added sodium methoxide (15 mmol) and refluxed for 7 h. The mixture so obtained was cooled to room temperature and excess of solvent was removed under vacuo. The obtained residue was extracted with ethyl acetate and dried over sodium sulphate to obtain the white coloured amorphous product **4** with 62 % yield. Mp: 292-294 °C; ^1H -NMR (CDCl_3): δ 1.45 (s, 9H, Boc-CH₃), 1.62 (m, 2H, Piperidine-H), 1.88 (m, 2H, Piperidine-H), 2.2 (m, 2H, Piperidine-H), 2.4 (m, 2H, Piperidine-H), 2.7 (s, 3H, N-CH₃), 3.12 (s, 3H, OCH₃), 3.79 (m, 2H, methylene-H), 7.21 (m, 1H, Ar-H), 7.80 (m, 1H, Ar-H); Anal. Calcd for C₁₉H₂₇N₃O₄; C, 63.14; H, 7.53; N, 11.63; O, 17.71; MS (ESI) m/z: 361.2.

Synthesis of 1-(6-methoxybenzo[d]oxazol-2-yl)-N-methylpiperidin-4-amine 5

Compound **4** (10 mmol) was taken in tetrahydrofuran (100 mL) and mixed to form a uniform mixture, to this was added potassium iodide (10 mmol) and refluxed for 12 h. The mixture so obtained was cooled to room temperature and excess of solvent was removed under vacuo. The obtained residue was treated with trifluoroacetic acid in dichloroethane to obtain deprotected amine **5** with 71 % yield. Mp: 167-169 °C; ^1H -NMR (CDCl_3): δ 1.11 (m, 2H, Piperidine-H), 1.34 (m, 2H, Piperidine-H), 2.1 (m, 2H, Piperidine-H), 2.6 (m, 2H, Piperidine-H), 2.7 (s, 3H, N-CH₃), 3.12 (s, 3H, OCH₃), 3.79 (m, 2H, methylene-H), 7.25 (m, 1H, Ar-H), 7.88 (m, 1H, Ar-H), 8.4 (s, 1H, NH); Anal. Calcd for C₁₄H₁₉N₃O₂; C, 64.35; H, 7.33; N, 16.08; O, 12.25; MS (ESI) m/z: 261.15.

General procedure for the synthesis of various N-substituted benzyl-1-(6-methoxybenzo[d]oxazol-2-yl)-N-methylpiperidin-4-amine 6a-l

In the last step compound **5** (0.2 mmol) was added to DMF (10 mL), to this solution anhydrous potassium carbonate (0.30 mmol) followed by various substituted benzyl chloride (0.2 mmol). This solution was stirred for 4 h at room temperature, on completion of reaction on the TLC water was added. The compound was extracted with DCM and dried over sodium sulphate, this was dried in vacuo to provide with final derivatives **6a-l**.

N-Benzyl-1-(6-methoxybenzo[d]oxazol-2-yl)-N-methylpiperidin-4-amine 6a

59 % yield. Mp: 157-159 °C; ^1H -NMR (CDCl_3): δ 1.43 (m, 2H, Piperidine-H), 1.68 (m, 2H, Piperidine-H), 2.65 (s,

3H, N-CH₃), 2.69 (m, 1H, Piperidine-H), 2.7 (m, 2H, Piperidine-H), 2.8 (m, 2H, Piperidine-H), 3.62 (s, 2H, methylene-H), 3.73 (s, 3H, OCH₃), 7.06 (m, 2H, Ar-H), 7.1 (m, 1H, Ar-H), 7.2 (m, 2H, Ar-H), 7.25 (m, 1H, Benzoxazolyl-H), 7.88 (m, 2H, Benzoxazolyl-H); ¹³C NMR spectral data (25.15 MHz, CDCl₃): δ 136.5 (C-2), 143.4 (C-4), 149.9 (C-5), 95.5 (C-6), 158.3 (C-7), 112.9 (C-8), 119.7 (C-9), 49.6 (C-11), 31.8 (C-12), 51.3 (C-13), 31.8 (C-14), 49.6 (C-15), 55.5 (C-17), 42.7 (C-19), 57.5 (C-20), 136.7 (C-21), 128.0 (C-22, C-26), 128.6 (C-23, C-25), 128.9 (C-24) Anal. Calcd for C₂₁H₂₅N₃O₂; C, 71.77; H, 7.17; N, 11.96; O, 9.10; MS (ESI) m/z: 351.1947.

N-(4-chlorobenzyl)-1-(6-methoxybenzo[d]oxazol-2-yl)-N-methylpiperidin-4-amine **6b**

51 % yield. Mp: 151-153 °C; ¹H-NMR (CDCl₃): δ 1.44 (m, 2H, Piperidine-H), 1.67 (m, 2H, Piperidine-H), 2.67 (s, 3H, N-CH₃), 2.68 (m, 1H, Piperidine-H), 2.75 (m, 2H, Piperidine-H), 2.82 (m, 2H, Piperidine-H), 3.63 (s, 2H, methylene-H), 3.75 (s, 3H, OCH₃), 7.08 (m, 2H, Ar-H), 7.29 (m, 2H, Ar-H), 7.42 (m, 1H, Benzoxazolyl-H), 7.88 (m, 2H, Benzoxazolyl-H); ¹³C NMR spectral data (25.15 MHz, CDCl₃): δ 136.5 (C-2), 143.4 (C-4), 149.2 (C-5), 95.9 (C-6), 158.4 (C-7), 112.4 (C-8), 119.7 (C-9), 49.3 (C-11), 31.5 (C-12), 51.3 (C-13), 31.8 (C-14), 49.7 (C-15), 55.7 (C-17), 42.4 (C-19), 57.3 (C-20), 136.5 (C-21), 129.6 (C-22, C-26), 131.8 (C-23, C-25), 124.2 (C-24) Anal. Calcd for C₂₁H₂₄ClN₃O₂; C, 65.36; H, 6.27; Cl, 9.19; N, 10.89; O, 8.29; MS (ESI) m/z: 385.1557.

N-(4-bromobenzyl)-1-(6-methoxybenzo[d]oxazol-2-yl)-N-methylpiperidin-4-amine **6c**

55 % yield. Mp: 154-156 °C; ¹H-NMR (CDCl₃): δ 1.39 (m, 2H, Piperidine-H), 1.59 (m, 2H, Piperidine-H), 2.59 (s, 3H, N-CH₃), 2.68 (m, 1H, Piperidine-H), 2.75 (m, 2H, Piperidine-H), 2.82 (m, 2H, Piperidine-H), 3.63 (s, 2H, methylene-H), 3.75 (s, 3H, OCH₃), 7.11 (m, 2H, Ar-H), 7.30 (m, 2H, Ar-H), 7.42 (m, 1H, Benzoxazolyl-H), 7.88 (m, 2H, Benzoxazolyl-H); ¹³C NMR spectral data (25.15 MHz, CDCl₃): δ 136.5 (C-2), 143.4 (C-4), 149.2 (C-5), 95.9 (C-6), 158.4 (C-7), 112.4 (C-8), 119.7 (C-9), 49.3 (C-11), 31.5 (C-12), 51.3 (C-13), 31.8 (C-14), 49.7 (C-15), 55.7 (C-17), 42.4 (C-19), 57.3 (C-20), 136.5 (C-21), 129.6 (C-22, C-26), 131.8 (C-23, C-25), 124.2 (C-24); Anal. Calcd for C₂₁H₂₄BrN₃O₂; C, 58.61; H, 5.62; Br, 18.57; N, 9.76; O, 7.44; MS (ESI) m/z: 429.1052

N-(4-fluorobenzyl)-1-(6-methoxybenzo[d]oxazol-2-yl)-N-methylpiperidin-4-amine **6d**

61 % yield. Mp: 150-152 °C; ¹H-NMR (CDCl₃): δ 1.43 (m, 2H, Piperidine-H), 1.66 (m, 2H, Piperidine-H), 2.67 (s, 3H, N-CH₃), 2.68 (m, 1H, Piperidine-H), 2.75 (m, 2H, Piperidine-H), 2.82 (m, 2H, Piperidine-H), 3.63 (s, 2H, methylene-H), 3.75 (s, 3H, OCH₃), 7.09 (m, 2H, Ar-H), 7.28 (m, 2H, Ar-H), 7.42 (m, 1H,

Benzoxazolyl-H), 7.88 (m, 2H, Benzoxazolyl-H); ¹³C NMR spectral data (25.15 MHz, CDCl₃): δ 136.1 (C-2), 143.9 (C-4), 149.3 (C-5), 95.5 (C-6), 158.3 (C-7), 112.6 (C-8), 119.5 (C-9), 49.4 (C-11), 31.4 (C-12), 51.6 (C-13), 31.5 (C-14), 49.5 (C-15), 55.5 (C-17), 42.1 (C-19), 57.2 (C-20), 136.5 (C-21), 129.8 (C-22, C-26), 115 (C-23, C-25), 163.4 (C-24); Anal. Calcd for C₂₁H₂₄FN₃O₂; C, 68.27; H, 6.55; F, 5.14; N, 11.37; O, 8.66; MS (ESI) m/z: 369.1853.

N-(3,4-difluorobenzyl)-1-(6-methoxybenzo[d]oxazol-2-yl)-N-methylpiperidin-4-amine **6e**

65 % yield. Mp: 169-171 °C; ¹H-NMR (CDCl₃): δ 1.44 (m, 2H, Piperidine-H), 1.67 (m, 2H, Piperidine-H), 2.67 (s, 3H, N-CH₃), 2.68 (m, 1H, Piperidine-H), 2.75 (m, 2H, Piperidine-H), 2.82 (m, 2H, Piperidine-H), 3.63 (s, 2H, methylene-H), 3.75 (s, 3H, OCH₃), 7.08 (m, 2H, Ar-H), 7.25 (m, 1H, Ar-H), 7.42 (m, 1H, Benzoxazolyl-H), 7.88 (m, 2H, Benzoxazolyl-H); ¹³C NMR spectral data (25.15 MHz, CDCl₃): δ 136.4 (C-2), 144.1 (C-4), 149.2 (C-5), 95.2 (C-6), 158.9 (C-7), 112.2 (C-8), 119.7 (C-9), 49.6 (C-11), 31.7 (C-12), 51.4 (C-13), 31.8 (C-14), 49.5 (C-15), 55.9 (C-17), 42.3 (C-19), 57.6 (C-20), 140.2 (C-21), 129.8 (C-22, C-26), 128.7 (C-22), 133.2 (C-23), 133.6 (C-24), 129.9 (C-25), 129.2 (C-26); Anal. Calcd for C₂₁H₂₃F₂N₃O₂; C, 65.10; H, 5.98; F, 9.81; N, 10.85; O, 8.26; MS (ESI) m/z: 387.1758.

N-(4-methylbenzene)-1-(6-methoxybenzo[d]oxazol-2-yl)-N-methylpiperidin-4-amine **6f**

61 % yield. Mp: 155-157 °C; ¹H-NMR (CDCl₃): δ 0.9 (s, 3H, CH₃), 1.42 (m, 2H, Piperidine-H), 1.66 (m, 2H, Piperidine-H), 2.66 (s, 3H, N-CH₃), 2.69 (m, 1H, Piperidine-H), 2.75 (m, 2H, Piperidine-H), 2.82 (m, 2H, Piperidine-H), 3.63 (s, 2H, methylene-H), 3.75 (s, 3H, OCH₃), 7.18 (m, 2H, Ar-H), 7.29 (m, 2H, Ar-H), 7.42 (m, 1H, Benzoxazolyl-H), 7.88 (m, 2H, Benzoxazolyl-H); ¹³C NMR spectral data (25.15 MHz, CDCl₃): δ 136.5 (C-2), 143.7 (C-4), 149.4 (C-5), 95.9 (C-6), 158.4 (C-7), 112.5 (C-8), 119.8 (C-9), 49.1 (C-11), 31.2 (C-12), 51.9 (C-13), 31.1 (C-14), 49.3 (C-15), 55.2 (C-17), 42.7 (C-19), 57.4 (C-20), 136.5 (C-21), 128.2 (C-22, C-26), 125.1 (C-23, C-25), 131.9 (C-24), 124.1 (C-27); Anal. Calcd for C₂₂H₂₇N₃O₂; C, 72.30; H, 7.45; N, 11.50; O, 8.76; MS (ESI) m/z: 365.2103.

N-(4-(trifluoromethyl)benzyl)-1-(6-methoxybenzo[d]oxazol-2-yl)-N-methylpiperidin-4-amine **6g**

70 % yield. Mp: 148-150 °C; ¹H-NMR (CDCl₃): δ 1.43 (m, 2H, Piperidine-H), 1.66 (m, 2H, Piperidine-H), 2.65 (s, 3H, N-CH₃), 2.69 (m, 1H, Piperidine-H), 2.74 (m, 2H, Piperidine-H), 2.82 (m, 2H, Piperidine-H), 3.63 (s, 2H, methylene-H), 3.75 (s, 3H, OCH₃), 7.18 (m, 2H, Ar-H), 7.29 (m, 2H, Ar-H), 7.42 (m, 1H, Benzoxazolyl-H), 7.88 (m, 2H, Benzoxazolyl-H); ¹³C NMR spectral data (25.15 MHz, CDCl₃): δ 136.2 (C-2), 143.5 (C-4), 149.1 (C-5), 95.2 (C-6), 158.7 (C-7), 112.3 (C-8), 119.4 (C-9), 49.3 (C-

11), 31.4(C-12), 51.7(C-13), 31.5(C-14), 49.4(C-15), 55.3(C-17), 42.2(C-19), 57.6 (C-20), 136.7(C-21), 128.5 (C-22, C-26), 128.3(C-23,C-25), 139.8 (C-24) , 21.3 (C-27); Anal. Calcd for $C_{22}H_{24}F_3N_3O_2$; C, 63.00; H, 5.77; F, 13.59; N, 10.02; O, 7.63; MS (ESI) m/z: 419.1821.

N-(4-*tert*-butylbenzyl)-1-(6-methoxybenzo[d]oxazol-2-yl)-*N*-methylpiperidin-4-amine **6h**

65 % yield. Mp: 168-170 °C; 1H -NMR ($CDCl_3$): δ 1.33 (s, 9H, CH_3), 1.43 (m, 2H, Piperidine-H), 1.66 (m, 2H, Piperidine-H), 2.65 (s, 3H, N- CH_3), 2.69 (m, 1H, Piperidine-H), 2.74 (m, 2H, Piperidine-H), 2.82 (m, 2H, Piperidine-H), 3.63 (s, 2H, methylene-H), 3.75 (s, 3H, OCH_3), 7.18 (m, 2H, Ar-H), 7.29 (m, 2H, Ar-H), 7.42 (m, 1H, Benzoxazolyl-H), 7.88 (m, 2H, Benzoxazolyl-H); ^{13}C NMR spectral data (25.15 MHz, $CDCl_3$): δ 136.5 (C-2), 143.3 (C-4), 149.8 (C-5), 95.3 (C-6), 158.3(C-7), 112.6(C-8), 119.2(C-9), 49.5(C-11), 31.5(C-12), 51.2(C-13), 31.5(C-14), 49.2(C-15), 55.8(C-17), 42.3(C-19), 57.7 (C-20), 136.8(C-21), 128.1 (C-22, C-26), 129.4(C-23,C-25), 144.5 (C-24) , 35.1 (C-27), 33.5(C-28), 29.1(C-29), 14.0(C-30); Anal. Calcd for $C_{25}H_{33}N_3O_2$; C, 73.68; H, 8.16; N, 10.31; O, 7.85; MS (ESI) m/z: 407.2573.

N-(4-nitrobenzyl)-1-(6-methoxybenzo[d]oxazol-2-yl)-*N*-methylpiperidin-4-amine **6i**

70 % yield. Mp: 177-179 °C; 1H -NMR ($CDCl_3$): δ 1.43 (m, 2H, Piperidine-H), 1.66 (m, 2H, Piperidine-H), 2.65 (s, 3H, N- CH_3), 2.69 (m, 1H, Piperidine-H), 2.74 (m, 2H, Piperidine-H), 2.82 (m, 2H, Piperidine-H), 3.63 (s, 2H, methylene-H), 3.75 (s, 3H, OCH_3), 7.18 (m, 2H, Ar-H), 7.29 (m, 2H, Ar-H), 7.42 (m, 1H, Benzoxazolyl-H), 7.88 (m, 2H, Benzoxazolyl-H); ^{13}C NMR spectral data (25.15 MHz, $CDCl_3$): δ 136.3 (C-2), 143.7 (C-4), 149.3 (C-5), 95.5 (C-6), 158.4(C-7), 112.7(C-8), 119.3(C-9), 49.5(C-11), 31.6(C-12), 51.8(C-13), 31.3(C-14), 49.4(C-15), 55.3(C-17), 42.9(C-19), 57.5(C-20), 136.7(C-21), 127.9 (C-22, C-26), 117.3(C-23,C-25), 140.5 (C-24) Anal. Calcd for $C_{21}H_{24}N_4O_4$; C, 63.62; H, 6.10; N, 14.13; O, 16.14; MS (ESI) m/z: 396.1798.

N-(2,5-dimethylbenzyl)-1-(6-methoxybenzo[d]oxazol-2-yl)-*N*-methylpiperidin-4-amine **6j**

55 % yield. Mp: 154-156 °C; 1H -NMR ($CDCl_3$): 1H -NMR ($CDCl_3$): δ 1.43 (m, 2H, Piperidine-H), 1.66 (m, 2H, Piperidine-H), 2.35 (s, 6H, CH_3), 2.65 (s, 3H, N- CH_3), 2.69 (m, 1H, Piperidine-H), 2.74 (m, 2H, Piperidine-H), 2.82 (m, 2H, Piperidine-H), 3.63 (s, 2H, methylene-H), 3.75 (s, 3H, OCH_3), 7.18 (m, 2H, Ar-H), 7.29 (m, 1H, Ar-H), 7.42 (m, 1H, Benzoxazolyl-H), 7.88 (m, 2H, Benzoxazolyl-H); ^{13}C NMR spectral data (25.15 MHz, $CDCl_3$): δ 136.6 (C-2), 143.8 (C-4), 149.2 (C-5), 95.4 (C-6), 158.5(C-7), 112.2(C-8), 119.8(C-9), 49.2(C-11), 31.3(C-12), 51.2(C-13), 31.6(C-14), 49.4(C-15), 55.2(C-17), 42.7(C-19), 53.7(C-20), 131.3(C-21), 129.2 (C-22),

127.8(C-26), 129.1(C-23),128.6(C-25), 139.4 (C-24), 18.7(C-25), 20.9(C-26); Anal. Calcd for $C_{23}H_{29}N_3O_2$; C, 72.79; H, 7.70; N, 11.07; O, 8.43; MS (ESI) m/z: 379.226.

N-(2,5-dimethoxybenzyl)-1-(6-methoxybenzo[d]oxazol-2-yl)-*N*-methylpiperidin-4-amine **6k**

67 % yield. Mp: 162-164 °C; 1H -NMR ($CDCl_3$): 1H -NMR ($CDCl_3$): δ 1.42 (m, 2H, Piperidine-H), 1.64 (m, 2H, Piperidine-H), 2.64 (s, 3H, N- CH_3), 2.69 (m, 1H, Piperidine-H), 2.73 (m, 2H, Piperidine-H), 2.83 (m, 2H, Piperidine-H), 3.63 (s, 2H, methylene-H), 3.72 (s, 6H, OCH_3), 3.80 (s, 3H, OCH_3), 7.18 (m, 2H, Ar-H), 7.29 (m, 1H, Ar-H), 7.42 (m, 1H, Benzoxazolyl-H), 7.89 (m, 2H, Benzoxazolyl-H); ^{13}C NMR spectral data (25.15 MHz, $CDCl_3$): δ 136.7 (C-2), 143.5 (C-4), 149.8 (C-5), 95.1 (C-6), 158.2(C-7), 112.7(C-8), 119.5(C-9), 49.3(C-11), 31.9(C-12), 51.4(C-13), 31.3(C-14), 49.3(C-15), 55.4(C-17), 42.2(C-19), 53.7(C-20), 127.7(C-21), 158.1 (C-22), 111.2(C-26), 112.3(C-23),157.7(C-25), 115.3 (C-24), 55.9(C-28), 55.5(C-30); Anal. Calcd for $C_{23}H_{29}N_3O_4$; C, 67.13; H, 7.10; N, 10.21; O, 15.55; MS (ESI) m/z: 411.2158.

N-(3,4,5-trimethoxybenzyl)-1-(6-methoxybenzo[d]oxazol-2-yl)-*N*-methylpiperidin-4-amine **6l**

72 % yield. Mp: 169-171 °C; 1H -NMR ($CDCl_3$): 1H -NMR ($CDCl_3$): δ 1.41 (m, 2H, Piperidine-H), 1.63 (m, 2H, Piperidine-H), 2.65 (s, 3H, N- CH_3), 2.68 (m, 1H, Piperidine-H), 2.73 (m, 2H, Piperidine-H), 2.84 (m, 2H, Piperidine-H), 3.61 (s, 2H, methylene-H), 3.75 (s, 9H, OCH_3), 3.80 (s, 3H, OCH_3), 7.19 (m, 1H, Ar-H), 7.27 (m, 1H, Ar-H), 7.41 (m, 1H, Benzoxazolyl-H), 7.90 (m, 2H, Benzoxazolyl-H); ^{13}C NMR spectral data (25.15 MHz, $CDCl_3$): δ 136.6 (C-2), 143.8 (C-4), 149.2 (C-5), 95.4 (C-6), 158.5(C-7), 112.2(C-8), 119.8(C-9), 49.2(C-11), 31.3(C-12), 51.2(C-13), 31.6(C-14), 49.4(C-15), 55.2(C-17), 42.7(C-19), 53.7(C-20), 142.7(C-21), 107.6 (C-22, C-26), 152.9(C-23,C-25),139.2 (C-24), 56.2(C-28, C32), 60.9(C-30); Anal. Calcd for $C_{24}H_{31}N_3O_5$; C, 65.29; H, 7.08; N, 9.52; O, 18.12; MS (ESI) m/z: 441.2264.

Molecular modelling and docking studies

Active site determination for Hsp70 and Hsp90

All the molecular modelling studies were carried out on the Schrodinger modelling suite (Version 15.3). The Heat Shock Proteins (Hsp) 70 and 90 were obtained from Brookhaven Protein Data Bank (RCSB-PDB). Initially the proteins crystals obtained from PDB were analysed and the active sites were determined. Hsp70 (PDB Id: 2XA4) and Hsp90 (PDB Id: 3FGR) were prepared in the protein preparation wizard of the Schrodinger suite for knocking out the bound ligands and cofactors, excess and nonessential water molecules were also removed. The proteins were optimised and

missing loops were filled and overlapping residues were corrected, finally both the target proteins were optimised for their conformational energies. These were further subjected to Q-site program to determine the active sites within the protein structure.¹⁴ The active sites were determined with regions such as hydrophobic and polar-regions as presented in figure 2.

Molecular docking studies

The compounds designed were subjected to energy minimisation by Ligprep module of the software. Glide module was employed for the molecular docking; compounds were docked in the grid designed in the module. The docking

was carried out with flexible ligands and the protein with the simulations made on the OPLS5.0 force field. All the results were based on 10 conformations for each molecule in the standard precision (SP) mode. The crystal structure Hsp70 (PDB Id: 2XA4) and Hsp90 (PDB Id: 3FGR) were both employed as the receptor with compounds **6a-I** as ligands.¹⁴ The results obtained from the simulation were obtained in the form of dock score and Glide energy, these values represent the minimum energies. Interactions between the ligand and residues were presented in the form of H-bond, van der Waals forces and the pi bonds. The results in the form of 3D and 2D representation were obtained for simplified understanding and presented as figure 2 and Fig 3

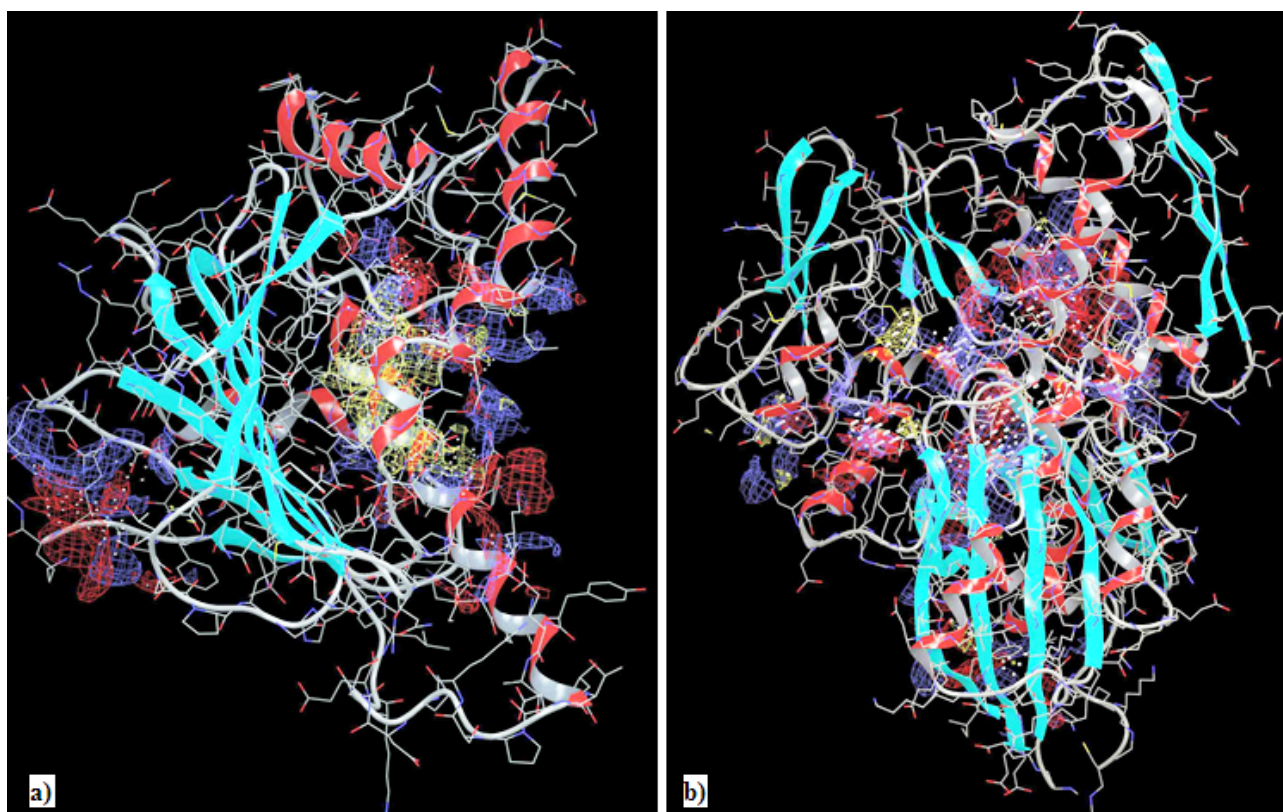


Figure 2: a) Active site determination in the Hsp70 (PDB Id: 2XA4); b) Active site determination in the Hsp90 (PDB Id: 3FGR)

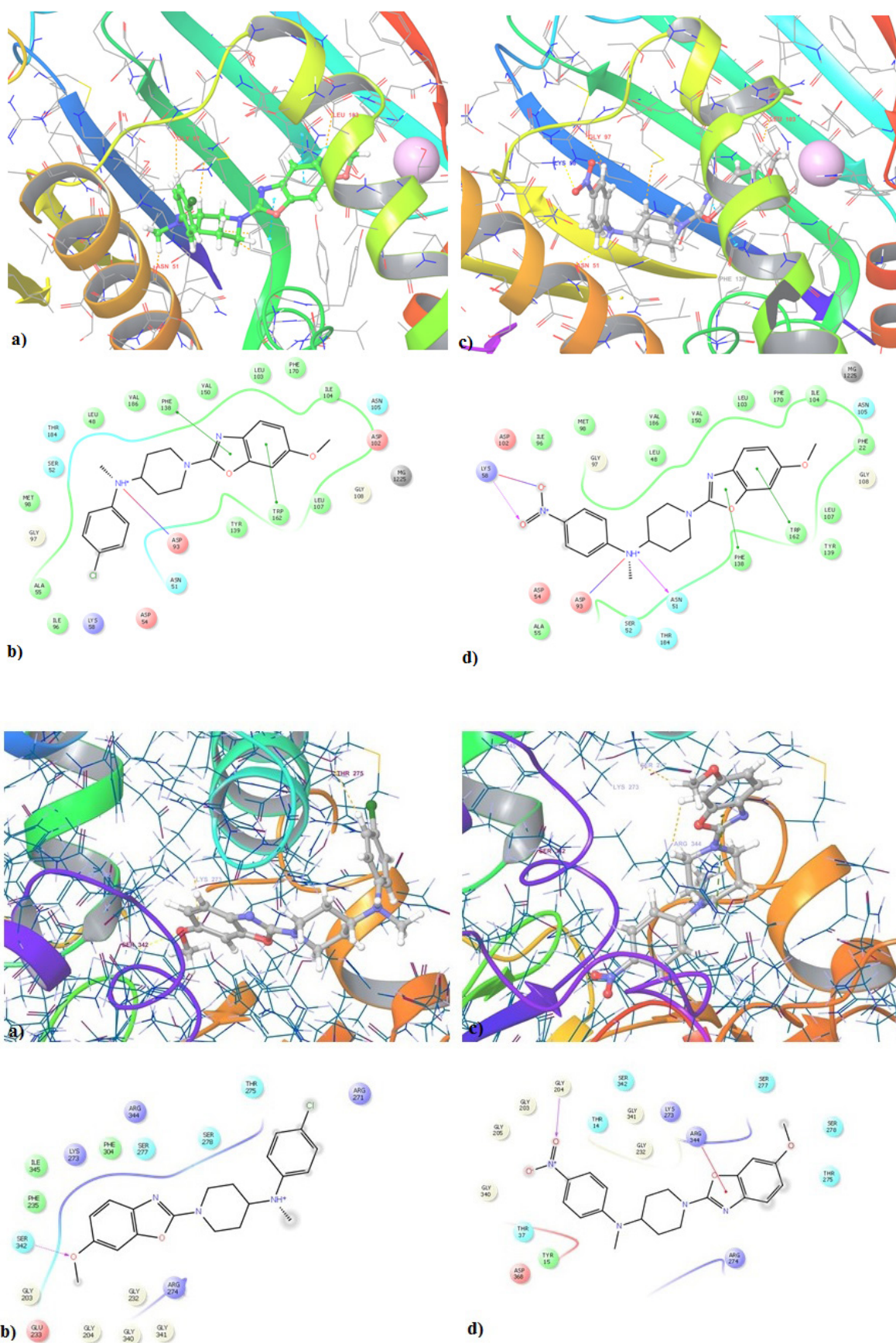


Figure 3: Molecular docking study on compounds from the series; a,b) 3D and 2D representation of compound 6bonHsp70 (PDB Id: 2XA4); c,d) 3D and 2D representation of compound 6ionHsp70 (PDB Id: 2XA4).

IC₅₀ for both the compounds are explained in figure 4A for **6b** and **6i**.

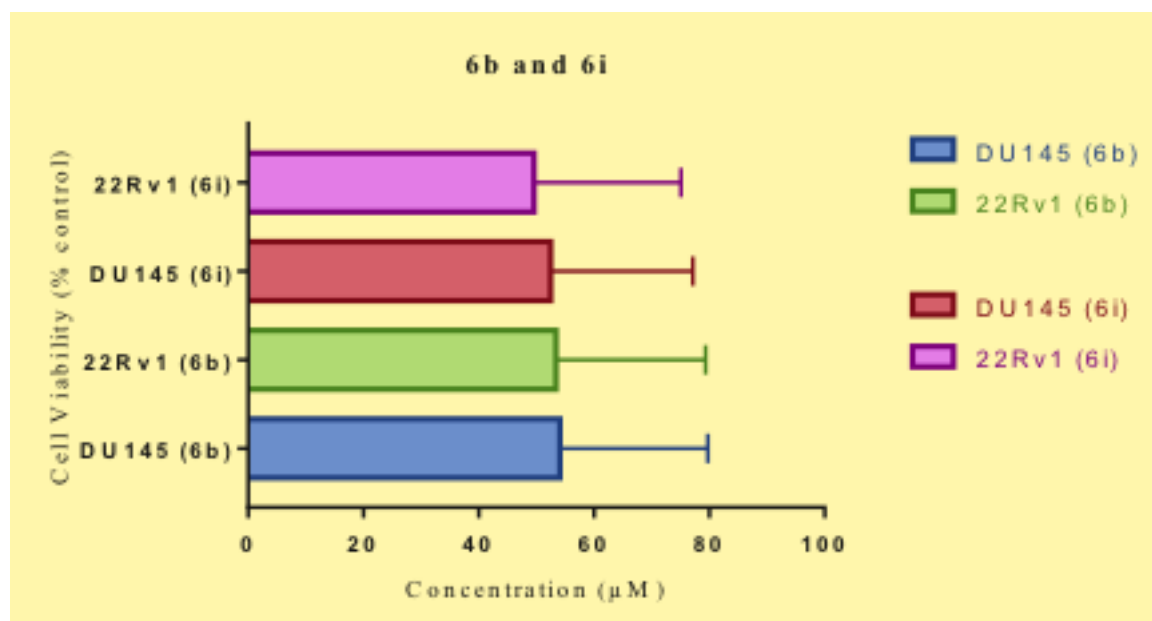


Figure 4. A) Molecular docking study on compounds from the series; a, b) 3D and 2D representation of compound **6b** on Hsp90 (PDB Id: 3FGR); c, d) 3D and 2D representation of compound **6i** on Hsp90 (PDB Id: 3FGR). B) Compound **6b** and **6i** inhibited overall cell viability (% relative to control) DU-145 and 22Rv1 (Cell lines for Prostate cancer). The cell viability was determined by plotting CCK-8 assay with increasing concentration of compounds for 24 hours.

Table 1: Antitumor activity of some novel hybrids of benzoxazole, the table represents concentration as a result of 50% loss of viable cells with respect to untreated cells (IC₅₀). The results were defined from dose-response curves for two different cell lines DU-145 and 22Rv1. It also presents the glide score and energies for different derivatives in front respective derivatives.

Sr. no.	Compd no.	IC ₅₀ (μM)		Dock score			
		DU145	22Rv1	Hsp 70		Hsp 90	
				Dock Score	Glide Energy	Dock Score	Glide Energy
1	6a	28.91	27.51	-8.0251	-46.3115	-5.07476	-34.435
2	6b	20.10	19.41	-8.12051	-46.1657	-5.08201	-49.2727
3	6c	>100	48.12	-7.76383	-48.4989	-4.41685	-43.5636
4	6d	47.45	44.94	-7.91281	-46.2466	-5.09304	-39.2727
5	6e	29.35	45.21	-8.02916	-49.2831	-5.34989	-46.6876
6	6f	>100	43.33	-8.02665	-46.5599	-3.7217	-35.7962
7	6g	45.24	35.55	-7.85481	-48.8817	-3.39334	-35.8336
8	6h	41.42	39.92	-7.51404	-46.4911	-3.51714	-38.4987
9	6i	20.5	19.00	-8.84901	-49.7121	-5.34706	-46.6326
10	6j	>100	43.36	-7.70333	-45.1409	-4.43773	-32.1075
11	6k	>100	44.33	-7.32014	-46.7713	-4.98797	-37.4438
12	6l	47.10	44.89	-7.63666	-49.3863	-3.6077	-39.2424
13	Std Enzalutamide	28.22	31.72	-7.344	-46.7819	-5.09304	-39.2727

In vitro screening

Cell Culture

The cell lines DU-145 and 22Rv1 were obtained from the ATCC, USA; DU145, 22Rv1 and RWPE-1 were preserved at -80 °C according to the standard protocol. These were further cultured in Dulbecco's Eagle medium modified and supplemented, it contains fetal Bovine Serum (10%) along with penicillin-100 U/mL and streptomycin-0.1 mg/mL as per the requirement. Before use these cell lines were kept for incubation at 37 °C with controlled humidity and 5 % carbon dioxide.

Oncotest assay method for antiproliferative activity^{15,16}

Oncotest's monolayer assay was developed for testing the antiproliferative activity of newly developed compounds was found to be the most accurate and suitable method for evaluation of these compounds. This method was found to be highly accurate and specific for testing of compounds on prostate cancer cell lines. The widely used anticancer drug enzalutamide was used as standard drug on both the cell lines along with other test compounds. This assay is based on the propidium iodide method, the test compounds are observed after four days of treatment. All the derivative **6a-1** were tested for ten dilutions with half log increasing to the 100 1M. The readings were taken in triplicates and observations were presented by nonlinear regression analysis as IC₅₀ in table 1 forming the results. The detailed experimental protocol is provided with the supplementary material available online.

Antitumor Activity by CCK-8 Assay¹⁷

The CCK-8 assay stands for Cell Counting Kit 8 (CCK 8) which was developed by Dojindo Corporation in Japan. It is a ready to use kit to determine the extent of proliferation by both cell lines DU-145 and 22Rv1 the selected compounds **6b** and **6i**. (Figure-4B). Three replicates for each of the compound was carried out and the cell growth inhibition was calculated as the following formula;

$$\text{Growth inhibition} = \frac{1 - OD \text{ of treated cells}}{OD \text{ of control cells}} \times 100\%$$

Surface Plasmon Resonance (SPR) Analysis¹⁸

All the SPR readings were obtained from optical biosensor Biacore T100 (GE Company) reported elsewhere. The human HSP70 (ADI-ESP-550-F [200 µg], Enzo Life Sciences) was immobilized on a CM5 sensor chip (GE) using 10 mmol/PBS at pH 5.0. The binding experiments were carried out at 25 °C using a flow rate of 30 mL/L at 90 s of

monitoring for association and 120 s of monitoring for dissociation of the ligand and substrate. All studies were performed with eight dilutions as 0.003, 0.01, 0.03, 0.1, 0.3, 1, 3, and 10 mM, with all of these prepared with 5% DMSO in PBS along with it 5% DMSO in PBS was used as a running buffer and blank control. The data was delivered using equilibrium dissociation constant (K_d value). This was calculated following equation as:

$$\text{Response} = \text{Conc} \times R_{\text{max}} / [\text{conc} + K_d] + \text{offset}$$

Table 2: The inhibitor binding of Heat Shock Protein 70 (Hsp70) by kinetic characterisation*

Sr. No.	Name	K _d (mol/L)
1	6b	2.030 × 10 ⁻⁶
2	6i	2.421 × 10 ⁻⁶
3	Std	2.066 × 10 ⁻⁶
* Determination of kinetic rate constants and affinities.		

RESULTS AND DISCUSSION

Prostate cancer has been treated with the help of chemotherapy and surgery, but rise in incidences of drug resistance in prostate carcinoma have inclined the treatment more towards surgery. This lead to need for newer and better drug molecules to treat the disease and hence forth discovery of newer targets. Hsp70 and Hsp90 has been studied for their role in the prostate carcinoma, it was found that there is an elevation and elevated expression of these proteins in the patients suffering from cancer. Further it was established that upregulation of these to heat shock proteins plays a significant role in cancer progression and hence are good targets for treatment of prostate cancer. Herein we have describe design and development of few novel inhibitors of Hsp70 and Hsp90 for their application towards treatment of prostate cancer. The design of new molecules is based on the structural features like the fused aromatic nucleus, cyclic aliphatic group and halogen substitutions on the aromatic ring. These features are found to be common in many anticancer agents with use in treatment of prostate cancer. On the basis of hybrid approach we designed a series of derivatives with a fused aromatic nucleus the benzoxazole, cyclic aliphatic group the piperidine linker and aromatic ring with various substitutions.

The synthesis was carried out in four steps, first step involved substitution of piperidine ring **2** on the 2-chloro benzoxazole **1** leading to formation of the tert-butyl 1-(6-chlorobenzo[d]oxazol-2-yl)piperidin-4-yl(methyl)carbamate **3**, this was further treated with sodium methoxide in methanol to obtain tert-butyl 1-(6-methoxybenzo[d]oxazol-2-yl)piperidin-4-yl(methyl) carbamate **4**. This was de-

protected by removing Boc to yield 1-(6-methoxybenzo[d]oxazol-2-yl)-N-methylpiperidin-4-amine **5**. In the final step various phenyl chlorides were substituted to yield corresponding derivatives as N-substituted benzyl-1-(6-methoxybenzo[d]oxazol-2-yl)-N-methylpiperidin-4-amine **6a-l**. These compounds were characterised for their structural confirmation and molecular structures were established.

To study the biological efficacy of these compounds on cotest assay method was performed on two cell lines DU145 and 22Rv1. The results obtained from this assay shows that two compounds **6b** and **6i** have good IC_{50} values (Table 1). Compound **6b** had IC_{50} of 20.10 and 19.41 for the DU145 and 22Rv1 respectively, while **6i** had IC_{50} of 20.5 and 19.00 for DU145 and 22Rv1 respectively, this is quite satisfactory compared to the standard drug enzalutamide. In the subsequent evaluation these compounds were studied on the basis of CCK-8 assay for the two cell lines DU145 and 22Rv1 and cell viability was studied. It was found that compound **6i** inhibits the 22Rv1 more efficiently than DU145 whereas **6b** has more inhibitory activity on the 22Rv1 (Fig 5). In order to understand the mechanism of action of these two molecules molecular docking studies were carried out. Two different receptors were obtained from the protein data bank, Hsp70 (PDB ID: 2XA4) and Hsp90 (PDB ID: 3FGR). Initially these two proteins were subjected to protein preparation leading to filling of missing loops and resolving the overlapping residues. After optimising these protein on the Schrodinger protein preparation wizard these were subjected to Qsite finder. Qsite determines the active site within the receptor which is important for determining the residues involved in interaction with the ligands (Figure 2). In case of Hsp70 it was found that residues Leu40, Val186, Asp93, Trp162, Phe138, Ser52, Leu103, Asn51 and Gly100 form the active site and consist of a cavity within the receptor. In Hsp90 the active site was formed of Ser342, Phe235, Ile345, Lys273, Ser277, Arg274, Ser278, Glu233 and Arg344.

The molecular docking was carried out for the two compounds **6b** and **6i** in these active sites of Hsp70 and Hsp90 with the help of Glide module of Schrodinger software version 9.5. The compounds were docked on both the receptors and scores were obtained for their binding glide energy and dock score. The results are shown in table 1, it can be seen that both compounds have performed well in the docking studies. Compound **6b** has good dock score of -8.12 and Glide score of -46.16 for Hsp70, for Hsp90 it had dock score of -5.08 and Glide energy of -49.27. Compound **6i** has good dock score of -8.84 and Glide score of -49.71 for Hsp70, for Hsp90 it had dock score of -5.34 and Glide energy of -46.63. Figure 3 shows the three dimensional and two dimensional images of the compounds **6b** and **6i** interacting with Hsp70. It is clearly observed that the compound **6b**

forms hydrogen bond with the amine group of the linker and Asp93, two pi bond interactions are observed between the Phe138 and Trp162 with the benzoxazole nucleus. Compound **6i** interacts with the receptor forming hydrogen bonds with Asn51 and Asp93, pi bond between Phe138 and Trp162, hydrogen and salt bridge with the Lys58. Figure 4 shows the three dimensional and two dimensional images of the compounds **6b** and **6i** interacting with Hsp90. It is observed that **6b** forms an hydrogen bond with Ser342 and **6i** forms a pi bond with the Arg344. These results suggest that compound **6b** and **6i** have high affinity for the Hsp70 compared to Hsp90.

In order to further investigate the efficacy of compound **6b** and **6i** we carried out the drug binding assay based on the surface plasmon resonance (SPR) principle on the Hsp70. The experiment was carried out on the human Hsp70 and the association and disassociation of the ligand with the substrate was determined (Table 2). The dissociation constant (K_d) was calculated and it was found that compound **6b** had $K_d = 2.030 \times 10^{-6}$ and **6i** had $K_d = 2.421 \times 10^{-6}$ compared to the standard with $K_d = 2.066 \times 10^{-6}$. This indicates that compound **6b** and **6i** are having comparable affinity towards the Hsp90.

CONCLUSION

Present study provides with two novel molecules **6b** and **6i**, these have good inhibitory activity against the Hsp70 and Hsp90, but it is more specific for the inhibition of the Hsp70. The results obtained and the observation make it clear that these compounds have good in vitro activity against prostate cancer. These compounds can be further evaluated for their in vivo activity against the prostate cancer and developed into lead molecules.

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